



INVISIBLE ENEMIES IN THE HUMAN BODY: BIOLOGICAL CHARACTERISTICS OF PARASITIC PROTOZOA

Kodirjonov Jakhongir Komilovich

Tashkent Medical Academy, Termiz Branch Faculty of General Medicine, Faculty 2 1st Year
Student

jqodirjonov39@gmail.com

Kungirotova Anorxol Inoyatovna

Assistant, Department of Medical Biology and Histology

ABSTRACT

Infections caused by parasitic protozoa remain a significant global public health challenge. This study analyzes the ultrastructural modifications, metabolic dependency, and immune evasion strategies of obligate intracellular pathogens, including *Plasmodium*, *Toxoplasma*, *Leishmania*, and *Trypanosoma*, at the molecular level. As a result of evolutionary genome reduction, these parasites have lost several autonomous biochemical pathways, including purine biosynthesis, and exhibit a strong dependence on the host cell's metabolic resources. Their immune evasion mechanisms are not limited to passive concealment; rather, they involve molecular mimicry, antigenic variation, and modulation of the intracellular environment of macrophages, including pH and redox balance. Certain pathogens may also activate Toll-like receptor (TLR) signaling pathways, leading to dysregulated cytokine production that contributes to tissue damage and inflammatory imbalance. At the same time, the dependence of parasites on host metabolic and oxidative stress systems represents a key vulnerability and a potential therapeutic target. In particular, antioxidant systems such as iron superoxide dismutase (Fe-SOD) and membrane transport proteins (ENTs) are considered promising targets for future drug development. Modern therapeutic strategies should therefore focus not only on direct parasite elimination but also on disrupting their metabolic supply chains.

Keywords: Parasitic protozoa, molecular mimicry, antigenic variation, metabolic dependency, immune modulation, cytokine response, parasitophorous vacuole, therapeutic targets.

INTRODUCTION

Endemic infections caused by parasitic protozoa such as *Plasmodium*, *Leishmania*, and *Trypanosoma* are responsible for millions of deaths annually worldwide (WHO, 2023). Despite advances in modern pharmacological interventions and molecular diagnostics, the persistent failure to interrupt transmission chains is not primarily attributable to drug shortages, but rather to the extraordinary biological and genetic plasticity of these microscopic pathogens (Hotez et al., 2020). Within the framework of the evolutionary "Red Queen" dynamics, protozoa that have transitioned to obligate parasitism have undergone radical genome reduction (Brockhurst et al., 2014). The loss of autonomous metabolic pathways has rendered them highly dependent on host-derived resources; however, this dependency is compensated by highly refined regulatory systems governing phenotypic adaptation and gene expression control (Lukeš et al., 2014). Such adaptations are not merely survival strategies but represent sophisticated mechanisms for the molecular-level exploitation of host cellular resources. Current research and clinical protocols are often focused on the macroscopic outcomes of pathogenesis, while the fundamental ultrastructural and biochemical mechanisms of invasion remain incompletely understood (McConville et al., 2015). We hypothesize that the evolutionary success of parasitic protozoa in the human host is not solely due to passive evasion of immune surveillance, but also to their ability to actively reprogram host metabolism through specialized organelle-mediated mechanisms. If primitive parasites have relinquished most autonomous biosynthetic capabilities to ensure survival, the question arises as to how they successfully invade and persist within the highly complex and hostile environment of human cells, overcoming their defensive barriers. The key to this



paradox does not lie in macroscopic symptoms but in the initial interaction between pathogen and host cell, during which the parasite does not merely enter the cell; rather, it begins to deconstruct and appropriate the cellular architecture using its unique ultrastructural machinery. It is precisely this process—deconstruction of host cellular architecture—that is mediated by the parasite’s highly specialized ultrastructural apparatus.

Cellular and Molecular Adaptations of Parasitic Protozoa

The fundamental challenge of intracellular parasitism is not the traversal of the host plasma membrane, but rather ensuring host cell survival during invasion. Premature host cell death would eliminate the parasite’s ecological niche before establishment. In apicomplexan parasites such as *Plasmodium* and *Toxoplasma*, this function is mediated by the apical complex. This structure is not merely a membrane-penetrating device; rather, it operates as a highly precise molecular injection system that delivers microneme and rhoptry proteins in a tightly regulated manner (Cowman et al., 2017). During invasion, the parasite does not rupture the host membrane. Instead, it invaginates the membrane in a controlled manner through secreted effector proteins, forming a parasitophorous vacuole (PV). This compartment acts as a highly selective, non-fusogenic “protective capsule” that shields the parasite from lysosomal degradation and acidic hydrolysis (Sibley, 2011). The host membrane remains structurally intact, while immune recognition is effectively minimized due to the absence of overt damage-associated signals. In trypanosomatid parasites, a key ultrastructural adaptation is the kinetoplast. Evolution has condensed their mitochondrial genome into a dense and highly organized network of kinetoplast DNA (kDNA). During transmission from insect vectors to the human bloodstream, these parasites undergo a rapid and coordinated remodeling of their respiratory metabolism mediated by this structure (Lukeš et al., 2014). In oxygen-rich environments, oxidative phosphorylation is downregulated almost immediately, and the organism shifts to a predominantly glycolytic energy production system. This represents not a simple metabolic adjustment but an extreme form of bioenergetic reprogramming. However, successful intracellular localization and immune evasion constitute only the initial phase of survival. Once safely enclosed within the parasitophorous vacuole, the parasite faces a critical constraint: its highly reduced genome lacks the capacity for autonomous biosynthesis of essential metabolites. Having relinquished most independent metabolic pathways, the parasite rapidly transitions to a second, more aggressive phase of infection. It establishes specialized molecular interfaces with the host cell’s metabolic core, effectively extracting nutrients and resources in a highly efficient and continuous manner.

Antigenic Variation and Molecular Mimicry

Successful exploitation of host resources does not guarantee long-term survival; the pathogen must also evade continuous immune surveillance. A direct biological confrontation with the immune system typically results in rapid elimination of the microbe. Therefore, parasitic protozoa adopt a state of near-complete molecular camouflage. One of the most effective strategies is antigenic variation. *Trypanosoma brucei* possesses a genome encoding more than 2,000 variant surface glycoprotein (VSG) genes, although only a single gene is expressed at any given time (Horn, 2014). Before macrophages and B lymphocytes can mount a targeted antibody response against a specific VSG variant, the parasite undergoes transcriptional switching, replacing its entire surface coat with a novel antigenic profile. This process functions analogously to an adaptive algorithm that continuously changes a vehicle’s license plate and appearance at every checkpoint. As a result, immune recognition is persistently delayed, and infection progresses in a characteristic wave-like pattern. Similarly, *Plasmodium falciparum* employs the *var* gene family to express PfEMP1 proteins on the surface of infected erythrocytes (Scherf et al., 2008). These proteins undergo frequent interchromosomal recombination, leading to continuous remodeling of their molecular structure. This mechanism not



only enables escape from antibody-mediated clearance but also facilitates cytoadherence of infected erythrocytes to vascular endothelium, thereby preventing their filtration and removal by the spleen. Beyond antigenic variation, molecular mimicry represents a more passive yet equally effective immune evasion strategy. In this case, the parasite does not merely change its antigenic identity but instead structurally imitates host molecules. For example, *Trypanosoma cruzi* expresses surface epitopes that closely resemble antigens found in human cardiac and neural tissues (Cunha-Neto et al., 2006). As a consequence, the immune system misidentifies the pathogen as self-tissue, leading to impaired immune targeting. This immunological confusion can trigger deleterious autoimmune responses, in which the host immune system begins to damage its own cells rather than eliminating the pathogen. Within this disrupted immunological landscape, the parasite persists and proliferates, effectively exploiting the host's own defense mechanisms as a survival advantage. After evading immune surveillance, the pathogen rapidly switches off its passive hiding mode. Defense is replaced by aggressive destruction. The parasite activates its biochemical arsenal — a cascade of tissue-degrading specific enzymes and toxins. *Cysteine proteases (EhCP)* form the core of the pathogenesis of *Entamoeba histolytica* (McKerrow et al., 2006). These molecules are not simple digestive catalysts. They are high-precision “molecular scissors” that cut through extracellular matrix (ECM) proteins of the human body — collagen, fibronectin, and laminin — at their molecular bonds. Epithelial barriers are destroyed. Tissue architecture collapses. The parasite opens a blood-stained pathway toward the bloodstream. This mechanism becomes even more complex in the activity of *Trypanosoma cruzi*, which produces the protease cruzipain (Cazzulo et al., 1997). In addition to acting as a powerful “biological bulldozer” that invades tissues, this enzyme breaks down kininogens in the blood, artificially increasing bradykinin production. Vascular permeability becomes pathologically elevated, allowing the infection to spread throughout the body without restriction. Intracellular degradation ultimately results in massive intoxication.

CONCLUSION

The evolutionary success of protozoan parasites does not lie in their biological simplicity, but in their ability to identify and exploit fundamental weaknesses within human cells. The pathological chain from molecular invasion to systemic collapse can be synthesized into three core theses:

1. Logistical piracy

The pathogen deliberately abandons independent biochemical networks and compensates for its genetic deficiencies by exploiting the host's central bioenergetic system. This is not simple “nutritional dependence,” but parasitic piracy that hijacks the main supply lines. Metabolism becomes fully monopolized.

2. Immunological provocation

The pathogen does not merely evade the immune system — it actively redirects it against the host. The macrophage's acidic environment becomes a biological battlefield, ROS defenses are neutralized, and through TLR hyperactivation, a lethal “cytokine storm” is deliberately triggered. The organism effectively destroys itself.

3. Ultrastructural aggression

The apical complex and kinetoplast are not just tools for cell entry. They are highly precise “molecular hacking systems” that disrupt host cellular architecture from within and appropriate its entire informational code. The cellular system is compromised and reprogrammed. Fundamental medicine must no longer be distracted by peripheral symptoms. Therapeutic strategies should focus on cutting the invisible supply lines between host and pathogen and targeting the parasite's antioxidant “bio-armor.” This approach is not merely empirical drug discovery — it is a direct strike



against the parasite's central logistics. The true strength of the parasite does not lie in its genes. It lies within our own cells.

REFERENCES:

1. Brockhurst, M. A., Koskella, B., & MacLean, R. C. (2014). Running with the Red Queen: the role of biotic conflicts in evolution. *Nature Reviews Genetics*, 15(12), 790-802.
2. Bullen, H. E., Jia, Y., Yamaryo-Botté, Y., Bisio, H., Zhang, O., Jemelin, K. E., ... & Soldati-Favre, D. (2016). Phosphatidic acid-mediated signaling regulates microneme secretion in *Toxoplasma*. *Cell Host & Microbe*, 19(3), 349-360.
3. Campagnaro, G. D., Quintana, J. F., & Field, M. C. (2018). Trypanosomatid genomes: evolution, expression, and variation. *Current Opinion in Microbiology*, 46, 17-23.
4. Cazzulo, J. J., Stoka, V., & Turk, V. (1997). Cruzipain, the major cysteine proteinase from the protozoan parasite *Trypanosoma cruzi*. *Biological Chemistry*, 378(12), 1369-1376.
5. Coombs, G. H., Goldberg, D. E., Klemba, M., Berry, C., Kelley, J., & Mottram, J. C. (2001). The malaria protease falcipain-2. *Trends in Parasitology*, 17(11), 532-537.
6. Coppens, I. (2006). Contribution of host lipids to *Toxoplasma* pathogenesis. *Cellular Microbiology*, 8(1), 1-9.
7. Coppens, I., Sinai, A. P., & Joiner, K. A. (2022). Metabolic exploitation of host cells by *Toxoplasma gondii*. *Annual Review of Microbiology*, 76, 213-235.
8. Cowman, A. F., Tonkin, C. J., Tham, W. H., & Duraisingh, M. T. (2017). The molecular basis of erythrocyte invasion by malaria parasites. *Cell*, 167(3), 610-624.
9. Cunha-Neto, E., Coelho, V., Guilherme, L., Fiorelli, A., Stolf, N., & Kalil, J. (2006). Autoimmunity in Chagas' disease. *Cardiovascular Research*, 71(3), 425-433.
10. Descoteaux, A., & Turco, S. J. (1999). Glycoconjugates in *Leishmania* infectivity. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1455(2-3), 341-352.
11. Egan, T. J. (2008). Haemozoin formation. *Molecular and Biochemical Parasitology*, 157(2), 127-136.
12. El Bissati, K., Zufferey, R., Witola, W. H., Carter, N. S., Ullman, B., & Ben Mamoun, C. (2010). The plasma membrane purine transporter of *Leishmania donovani*. *Journal of Biological Chemistry*, 285(13), 9425-9433.
13. Gazzinelli, R. T., Kalantari, P., Fitzgerald, K. A., & Golenbock, D. T. (2014). Innate sensing of malaria parasites. *Nature Reviews Immunology*, 14(11), 744-757.
14. Gazzinelli, R. T., Oliveira, A. C., & Cunha-Neto, E. (2021). The paradox of oxidative stress and parasite survival in macrophages. *Immunological Reviews*, 301(1), 112-126.
15. Greig, N., Wyllie, S., Patterson, S., & Fairlamb, A. H. (2015). A comparative study of the role of superoxide dismutases in *Leishmania*. *Open Biology*, 5(2), 140224.
16. Horn, D. (2014). Antigenic variation in African trypanosomes. *Molecular and Biochemical Parasitology*, 195(2), 123-129.
17. Hotez, P. J., Fenwick, A., Ray, S. E., Hay, S. I., & Molyneux, D. H. (2020). "Forgotten people, forgotten diseases"—the neglected tropical diseases and their impact on global health and development. *PLoS Neglected Tropical Diseases*, 14(1), e0008000.
18. Hunter, C. A., & Sibley, L. D. (2012). Modulation of innate immunity by *Toxoplasma gondii* virulence effectors. *Nature Reviews Microbiology*, 10(11), 766-778.
19. Jackson, A. P., Allison, H. C., Barry, J. D., Field, M. C., & Hertz-Fowler, C. (2016). A cell-surface phylome for African trypanosomes. *PLoS Neglected Tropical Diseases*, 10(3), e0004118.
20. Landfear, S. M. (2011). Nutrient transport and pathogenesis in selected parasitic protozoa. *Eukaryotic Cell*, 10(4), 483-493.



21. Lukeš, J., Hashimi, H., & Zíková, A. (2014). Unexplained complexity of the mitochondrial genome and transcriptome in kinetoplastid flagellates. *Current Genetics*, 60(1), 21-27.
22. McConville, M. J., de Souza, D. P., Saunders, E., Likic, V. A., & Naderer, T. (2015). Living in a phagolysosome; metabolism of *Leishmania* amastigotes. *Trends in Parasitology*, 31(10), 456-465.
23. McKerrow, J. H., Caffrey, C., Kelly, B., Loke, P., & Sajid, M. (2006). Proteases in parasitic diseases. *Annual Review of Pathology: Mechanisms of Disease*, 1, 497-536.
24. Romano, J. D., Sonda, S., Bergbower, E., Smith, M. E., & Coppens, I. (2013). *Toxoplasma gondii* salvages sphingolipids from the host Golgi through the rerouting of selected Rab vesicles. *Molecular Biology of the Cell*, 24(12), 1974-1995.
25. Scherf, A., Lopez-Rubio, J. J., & Riviere, L. (2008). Antigenic variation in *Plasmodium falciparum*. *Annual Review of Microbiology*, 62, 445-470.
26. Sibley, L. D. (2011). Invasion and intracellular survival by protozoan parasites. *Immunological Reviews*, 240(1), 72-91.
27. World Health Organization. (2023). *World malaria report 2023*. Geneva: World Health Organization.